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Catalytic FeCl₃- or Yb(OTf)₃-mediated synthesis of substituted tetrahydrofurans and C-aryl glycosides from 1,4-diols[†]

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Abstract—A facile and mild protocol for the synthesis of substituted tetrahydrofurans, 2-deoxy C-aryl glycoside and C-aryl glycosides is described by use of 20 mol% FeCl₃ or Yb(OTf)₃ as acid catalysts. The benzylic carbocation generated undergoes intramolecular substitution by the oxygen nucleophile to result the tetrahydrofurans and C-aryl glycosides. © 2002 Elsevier Science Ltd. All rights reserved.

1. Introduction

The synthesis of substituted tetrahydrofurans,¹ C-aryl and 2-deoxy-C-aryl glycosides, structural features that are commonly encountered in a variety of natural products,²⁻⁴ has attracted considerable attention due to their significant biological properties. Even though several acidic reagents⁵⁻⁹ (mineral, Lewis and Bronsted) were reported for the intramolecular cyclo dehydration of 1,4-diols, most of these methods use the reagents in stoichiometric quantities, require elevated temperatures and longer reaction times to synthesize the tetrahydrofurans. Recent years have witnessed an upsurge in the synthesis of C-aryl glycosides^{10–13} owing to the significant biological and pharmacological properties of some of these compounds. Taking advantage of facile carbocation formation in the presence of a variety of Lewis acids such as $FeCl_3$,¹⁴ DIBAL-H¹⁵ and Yb(OTf)₃,^{16–19} we have recently reported methods in protection-deprotection chemistry. In continuation of our studies on the use of carbocation chemistry in organic synthesis, herein, we describe a facile FeCl₂-²⁰or Yb(OTf)₃-mediated protocol for the conversion of 1,4diols (Eq. (1)) into substituted tetrahydrofurans, 2deoxy sugar and C-aryl glycosides.



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2. Results and discussion

Initially (Table 1), on reaction with 20 mol% FeCl₃ in CH₂Cl₂ at room temperature, diol 1 underwent facile cyclisation in 45 min to afford the tetrahydrofuran 1a as a racemic mixture (71%). Similarly 1,4-diol 2 under the above reaction conditions gave the corresponding tetrahydrofuran 2a (77%) as a 2:1 (trans:cis) mixture. The C(5) proton signal for the *trans* isomer appeared at δ 4.98 as a triplet (J=8.37 Hz) while for the *cis* isomer, it resonated as a doublet by triplet (J=2.32, 7.44 Hz) at δ 4.84. The diol **3** afforded **3a** in 73% yield as a single isomer (*trans*), $[\alpha]_{\rm D} = -43.3$ (c 1.7, CHCl₃), while the diol 4 gave 2-deoxy sugar 4a in 81% yield (1:1). When diols 1-4 were treated with 20 mol% Yb(OTf)₃ in CH₂Cl₂ at room temperature the expected products 1a-4a were formed. However, the reaction times were longer and the yields were lower than those from reactions with FeCl₃ (Table 1).

Having successfully prepared the tetrahydrofurans and 2-deoxysugar derivative, the protocol was then extended to the synthesis of *C*-aryl glycosides. Accord-

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Table 1. 20 mol% FeCl₃- and Yb(OTf)₃-mediated cyclisation of 1,4-diols

S. No	Starting Material	Product	20 mol% FeCl ₃		20 mol% Yb(OTf) ₃	
			Time (min)	Yield in % (Ratio of Isomers)	Time (h)	Yield in % (Ratio of Isomers)
1.	С s 1 ^{OH} OH		45	71 racemic	2.5	61 racemic
2.	$ \begin{array}{c} \begin{array}{c} \begin{array}{c} \begin{array}{c} \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\$	\downarrow_{0} \downarrow_{0} Ph 2a	45	77 (2:1, trans:cis)	4.5	52 (2:1, trans:cis)
3.	HO H	$ \begin{array}{c} $	45	73 (100:0 <i>trans:cis</i>)	3	67 (100:0 trans:cis)
4.	HO 4 OCH _{3OH}	HO Ph H ₃ CO 4a	45	81 (1:1 <i>trans:cis</i>)	2	71 (1:1 trans:cis)

ingly, diol **5** (Table 2) on treatment with 20 mol% FeCl₃ gave the *C*-aryl glycoside **5a**, albeit in a very poor yield.

Our earlier experience with Yb(OTf)₃ prompted us to treat the diols 5 and 6 with 20 mol% Yb(OTf)₃ in dry CH₂Cl₂ at 40°C, to afford C-aryl glycosides 5a (74%) and **6a** (71%), respectively as 3:2 (β : α) mixture of inseparable isomers. 1,4-Diols 7, 8 and 9, prepared from diacetone-D-mannose, when subjected to cyclization under the above reaction conditions gave the C-aryl glycosides 7a (72%), $[\alpha]_D = 19.4$ (c 1.2, CHCl₃), **8a** (68%), $[\alpha]_D = 12.4$ (c 1.2, CHCl₃) and **9a** (63%), $[\alpha]_{\rm D} = -10.5$ (c 1.6, CHCl₃) respectively, as exclusive α -isomers. The C(1) proton signal for 7a appeared at δ 5.0 as a doublet (J=8.6 Hz), for 8a at δ 5.28 as a singlet²¹ and for **9a** the C(1)H resonated at δ 5.29 as a doublet (J = 7.8 Hz). Similarly, the 1,4-diol 10, prepared from D-ribose, furnished the expected β -isomer 10a in 68% yield as an exclusive product, $([\alpha]_{D})$ = -17.7 (c 0.9, CHCl₃).

The structures of compounds 7a and 9a as exclusive α -isomers were unambiguously confirmed from NOE studies (Fig. 1). In compound 7a, C(1)H and C(2)H show NOE enhancements with the *ortho* protons of the phenyl ring, while 9a shows an NOE enhancement for C(2)H with the lone *ortho* proton, thus confirming the assigned structures.

3. Conclusions

In summary, a facile and mild protocol for the synthesis of substituted tetrahydrofurans, 2-deoxy sugar and C-aryl glycosides is described using 20 mol% FeCl₃ or Yb(OTf)₃ as acid catalysts. This methodology would be of immense use for synthesizing chiral furans as well as C-aryl glycosides.

4. Experimental

4.1. Typical experimental procedure

To a solution of diol 1 (0.2 g, 1.16 mmol) in CH_2Cl_2 (5 mL), anhydrous FeCl₃ (0.038 g, 0.23 mmol) was added and the mixture was stirred at room temperature for 45 min. After completion of the reaction (TLC analysis), it was diluted with CH_2Cl_2 (10 mL) and washed with saturated aq. NaHCO₃ solution (10 mL), water (10 mL), brine (10 mL) and dried (Na₂SO₄). Evaporation of solvent and purification by column chromatography (Si-gel, 5% EtOAc in hexane) gave 1a in 71% yield as a syrup. A similar procedure was adopted for the Yb(OTf)₃ mediated synthesis of *C*-aryl glycosides.

4.2. Spectral data for selected compounds

Compound 3a: ¹H NMR: (400 MHz, CDCl₃, TMS, δ in ppm): 1.30 (s, 3H, -CH₃), 1.50 (s, 3H, -CH₃), 1.79–1.9

Table 2. Conversion of 1,4-diols into C-aryl glycosides using 20 mol% Yb(OTf)₃

S.No.	Starting Material	Product	Time (h)	Yield(%)	Ratio of Isomers
1.	BnO OHHO S BnO OBn	$B_{nO} \xrightarrow{\tilde{c}}_{BnO} \xrightarrow{\tilde{c}}_{\bar{O}Bn} + other isomer 5a$	6	74	3:2 (β:α)
2.	BnO OHHO BnO OBn	BnO BnO + other isomer 6a	4	71	3:2 (β:α)
3.			6	72	exclusive (α)
4.		7a	5	68	exclusive (α)
5.		$\begin{array}{c} & & & \\ & & & & \\ & & & \\ & & & \\ & & & & \\ & & & & \\ & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & &$	8	63	exclusive (α)
6.			5	68	exclusive (β)
			-		
		0 0 11 H		H	-F
	7a 9a				

Figure 1. NOE interactions in compounds 7a and 9a.

(m, 1H, H-6), 1.96–2.08 (m, 1H, H-6"), 2.18–2.26 (m, 1H, H-7), 2.32–2.42 (m, 1H, H-7'), 3.48 (s, 3H, -OCH₃), 3.79 (d, J=10.2 Hz, 1H, H-4), 4.00–4.06 (m, 1H, H-3), 4.38–4.46 (m, 1H, H-5), 4.52 (d, 1H, J=10.2 Hz, H-2), 4.98–5.04 (t, 1H, J=11.28 Hz, H-8), 5.86 (d, 1H, J=10.2 Hz, H-1), 7.18–7.34 (m, 5H, Ar-H); FAB-MS (m/z, %): 319 (M⁺–1, 13.8), 147 (72.6), 91 (56.9), 81 (100), 77 (31.9).

Compound 4a: ¹H NMR: (400 MHz, CDCl₃, TMS, δ in ppm): 1.8–2.01 and 2.42–2.68 (2m, 2H, H-2.2¹), 3.36 (s, 3H, -OCH₃), 3.4 (s, 3H, -OCH₃), 3.61–4.08 (m, 3H, H-4.5), 4.10–4.20 and 4.28–4.34 (2m, 1H, H-3), 5.00–5.05 and 5.10–5.18 (2m, 1H, H-1), 7.2–7.38 (m, 5H, Ar-H); EIMS (m/z, %): 208 (M⁺, 63.4), 207 (M⁺–1, 57.2), 177 (16.4), 124 (47.5), 77 (100).

Compound 5a: ¹H NMR: (400 MHz, CDCl₃, TMS, δ in ppm): 3.64–3.78 (m, 2H, H-5), 3.8–3.86 (m, 1H, H-4), 4.04–4.10 (m, 1H, H-3), 4.18–4.6 (m, 7H, benzylic CH₂, H-2), 5.04, 5.38 (2d, *J*=11.0 Hz, H-1, 1H (α : β =2:3), 6.88–7.32 (m, 15H, Ar-H); FAB-MS (*m*/*z*, %): 509 (M⁺+23, 9.7), 485 (M⁺–1, 2.7), 281 (36.1) 181 (6.9), 91 (100).

Compound 8a: ¹H NMR: (400 MHz, CDCl₃, TMS, δ in ppm): 1.28 (s, 3H, -CH₃), 1.36 (s, 3H, -CH₃), 1.41 (s, 3H, -CH₃), 1.55 (s, 3H, -CH₃), 3.82–3.86 (m, 1H, H-4), 4.02–4.14 (m, 2H, H-6), 4.38–4.44 (m, 1H, H-5), 4.78–4.82 (m, 1H, H-3), 5.00 (d, 1H, J=9.9 Hz, H-2), 5.28 (s, 1H, H-1), 6.94–6.98 (m, 1H, Ar-H), 7.24–7.28 (m, 2H, Ar-H); FAB-MS (m/z, %): 349 (M⁺+23, 16.5), 327 (M⁺+1, 32), 311 (48) 303 (18) 143 (75).

Compound 10a: ¹H NMR: (200 MHz, CDCl₃, TMS, δ in ppm): 0.11 (s, 6H, -CH₃), 0.98 (s, 9H, -CH₃), 1.34 (s, 3H, -CH₃), 1.45 (s, 3H, -CH₃), 3.74-3.88 (m, 6H, -OCH₃, H-5, 6), 4.16–4.22 (m, 1H, H-4), 4.68–4.74 (m, 1H, H-3), 4.89 (d, 1H, J=4.76 Hz, H-2), 5.12 (d, 1H, J=4.76 Hz, H-1), 6.83 (d, J=9.30 Hz, 2H, Ar-H), 7.24 (d, 2H, J=9.30 Hz, Ar-H). FAB-MS (m/z, %): 393 (M⁺-1, 4.6), 391 (M⁺-3, 12.5), 359 (27.7), 279 (11.1), 121 (100).

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